

CardiologyRounds™

www.cardiologyrounds.org

AS PRESENTED IN THE ROUNDS OF THE CARDIOVASCULAR DIVISION
OF BRIGHAM AND WOMEN'S HOSPITAL, BOSTON, MASSACHUSETTS

Glucose-insulin-potassium (GIK) for acute myocardial infarction: Mechanisms of action and current status

CARL S. APSTEIN, M.D.

Recent basic and clinical evidence suggests that glucose-insulin-potassium (GIK) may reduce mortality after myocardial infarction (MI). This issue of *Cardiology Rounds* reviews the metabolic mechanisms responsible for the protective effects of GIK and its potential benefits in the management of acute MI (AMI).

Cardiac energy metabolism

The normal human heart requires a huge amount of energy. Adenosine triphosphate (ATP) delivers the energy required for contraction, relaxation, and ion homeostasis. A normal heart weighing 300 g synthesizes and utilizes approximately 15 times its weight (5 kg) of ATP each day. However, the heart contains only about 750 mg of ATP and, therefore, this critical but small ATP pool completely "turns over," (ie, is utilized and resynthesized) every few seconds. Any reduction in ATP synthesis quickly results in systolic and diastolic dysfunction that is soon followed by irreversible injury.

The metabolic energy source for this large amount of ATP synthesis comes primarily from the uptake and oxidation of glucose and fatty acids (usually referred to as free fatty acids [FFAs] because they are not esterified). The heart is metabolically dependent on an adequate supply of these substrates and an abundant supply of oxygen for their oxidation.¹ Thus, the heart is as much a furnace as it is a pump; it oxidizes hydrocarbon fuel (glucose and FFAs) to obtain the energy necessary for its function. Like a furnace or a hybrid engine, the heart can run on either or both of these two energy sources. Under well-oxygenated conditions, there is no apparent advantage to either substrate and when both are available, the heart utilizes both. However, during ischemia, when oxygen availability is limited, glucose is a better substrate (the reasons are discussed in detail below). A summary of glucose and FFA metabolism is shown in Figure 1.

Effects of infarction and ischemia on cardiac energy metabolism

During an acute MI, the heart is composed of several regions with important metabolic differences. In the non-ischemic region, workload is increased to compensate for the loss of function in the ischemic region. To accomplish a greater workload, the non-ischemic region requires an increased rate of ATP synthesis and, therefore, a higher rate of substrate oxidation. However, in the ischemic region, ATP synthesis is markedly reduced due to the lack of oxygen and washout of metabolic products. Furthermore, the extent and consequences of the metabolic alterations depend importantly on the severity of the ischemic state.

During an acute MI, residual or collateral flow usually provides at least 10% of the normal perfusion to a significant portion of the ischemic myocardium.^{2,5} This decrease in perfusion markedly reduces ATP synthesis. However, this small amount of oxygen still supports a level of oxidative ATP synthesis that greatly exceeds that produced from anaerobic glycolysis.^{6,8} Moreover, a level that is 10% of normal perfusion appears to provide an adequate washout of tissue lactate and a concomitant increase in ATP synthesis by GIK.^{6,7} In sub-regions, with more severe ischemia, anaerobic glycolysis becomes a progressively more important source of energy for a limited amount of ATP, which may or may not suffice to support the most essential cellular functions. Thus, during an acute MI, a mixture of both oxidative and anaerobic metabolism supports the myocardium.

Glycogen is rapidly mobilized during ischemia and reduced glycogen concentrations impair force development, calcium release, and contractile function.⁹ Key intermediates of the Krebs cycle also become depleted and may further impair ATP synthesis.^{1,10} Overall, the ischemic myocardium is metabolically characterized by:

- decreased synthesis of ATP with resultant reduced levels of ATP and creatine phosphate (CrP), and a corresponding increase in adenosine diphosphate (ADP) and inorganic phosphate (Pi)
- intracellular acidosis secondary to the proton generation from ATP breakdown, production of lactate from any anaerobic glycolysis, and the reduced washout level of the ischemic state
- increased cellular levels of calcium and sodium as ion homeostasis deteriorates
- depleted levels of Krebs cycle intermediates.



BRIGHAM AND
WOMEN'S HOSPITAL



HARVARD
MEDICAL SCHOOL
TEACHING AFFILIATE

Cardiovascular Division (Clinical)

Michelle Albert, MD	Eldrin Lewis, MD
Elliott Antman, MD	James Liao, MD
Donald S. Baim, MD	Peter Libby, MD (<i>Division Chief</i>)
Kenneth Baughman, MD	Leonard Lilly, MD
Joshua Beckman, MD	Bernard Lown, MD
Charles M. Blatt, MD	William Maisel, MD
Eugene Braunwald, MD	Thomas Michel, MD, PhD
Christopher Cannon, MD	David Morrow, MD
Ming Hui Chen, MD	Karen Moulton, MD
Michael Chin, MD, PhD	Gilbert Mudge, MD
Mark Creager, MD	Anju Nohria, MD
Elazer Edelman, MD, PhD	Patrick O'Gara, MD
Andrew Eisenhauer, MD	Marc A. Pfeffer, MD, PhD (<i>Editor</i>)
Laurence Epstein, MD	Jorge Plutzky, MD
James Fang, MD	Jeffrey Popma, MD
Mark Feinberg, MD	Shmuel Ravid, MD
Daniel Forman, MD	Frederic Resnic, MD
Jonas Galper, MD, PhD	Paul Ridker, MD
Peter Ganz, MD	Thomas Rocco, MD
J. Michael Gaziano, MD	Campbell Rogers, MD
Marie Gerhard-Herman, MD	Maria Rupnick, MD, PhD
Robert Gugliano, MD	Arthur Sasahara, MD
Michael Givertz, MD	S. Dinakar Satti, MD
Samuel Z. Goldhaber, MD	Jay Schneider, MD
Thomas B. Graboys, MD	Christine Seidman, MD
Howard Hartley, MD	Andrew Selwyn, MD
Carolyn Ho, MD	Daniel Simon, MD
Mukesh Jain, MD	Laurence Sloss, MD
John Jarcho, MD	Kyoko Soejima, MD
Paula Johnson, MD	Regina Sohn, MD
Scott Kinlay, MD	Scott Solomon, MD
Jamil Kirdar, MD	Lynne Stevenson, MD
James Kirshenbaum, MD	William Stevenson, MD
Gideon Koren, MD	Peter Stone, MD
Richard Kuntz, MD	Michael Sweeney, MD
Raymond Kwong, MD	Frederick Welt, MD
Michael J. Landzberg, MD	Justina Wu, MD
Richard Lee, MD	

Brigham and Women's Hospital

Fax: (617) 732-5291 Website: www.heartdoc.org

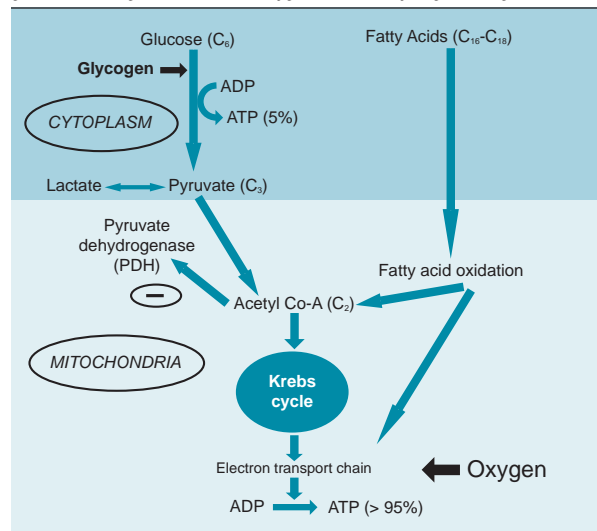
The editorial content of *Cardiology Rounds* is determined solely by the Cardiovascular Division of Brigham and Women's Hospital. This publication is made possible by an educational grant.

Cardiology Rounds is approved
by the Harvard Medical School
Department of Continuing Education
to offer continuing education credit

Figure 1. Comparison of glucose and FFA metabolism.

The lower half of the figure depicts the Krebs cycle and electron transport chain where oxidative ATP synthesis occurs. Both glucose and FFA metabolism result in the formation of the two carbon fragment, acetyl Co-A.

The upper half of the figure depicts the initial breakdown of glucose and FFA. The breakdown of glycogen and glucose to pyruvate occurs via the glycolytic pathway, where a small amount of ATP is synthesized. Under aerobic conditions, the pyruvate is converted to acetyl Co-A via the PDH reaction; under anaerobic conditions, the pyruvate is converted to lactate, which can diffuse out of the cell if adequate washout is present. The major FFAs are 16- and 18-carbon chains; these are broken down in successive intermediate reactions to two carbon fragments to generate the acetyl Co-A for the Krebs cycle. Under hypoxic cellular conditions, when acetyl Co-A is not actively utilized and removed in the Krebs cycle, partially metabolized FFA intermediates accumulate in the cytosol and have toxic effects due to their detergent-like properties (see text). The glucose and FFA pathways are mutually competitive. When FFA metabolism generates a high level of acetyl Co-A, the PDH reaction is inhibited, thereby decreasing the rate of flux through the glycolytic pathway. Conversely, a high level of plasma glucose and insulin, stimulates the glycolytic pathway and decreases myocyte FFA uptake. Because of differences in the two overall pathways, approximately 11% more oxygen is required to generate an equivalent amount of ATP from FFA than from glucose;¹⁶ thus, glucose is a more oxygen-efficient way of generating ATP.



FFA toxicity during MI

During an AMI, plasma FFA levels are increased in most patients secondary to the lipolytic action of endogenous or therapeutically-administered catecholamines and/or heparin.^{10a} High levels of plasma FFA increase myocyte FFA uptake and depress myocardial contractility, inhibit glycolytic flux, increase cyclic AMP levels, accumulate as intracellular toxic fatty acid derivatives, cause membrane damage and arrhythmias, and increase myocardial oxygen consumption without a concomitant increase in myocardial work (oxygen wasting effect of FFA).¹¹⁻¹⁴ Ischemia decreases the oxidation of FFA to carbon dioxide, resulting in increased intracellular accumulation of intermediary FFA metabolites such as acylcarnitine and acyl Co-A that have toxic effects. Acylcarnitine inhibits the sarcoplasmic reticular Ca^{2+} pump and the sarcolemmal Na^+/Ca^{2+} exchanger and Na^+ pump and can also activate Ca^{2+} channels and increase cyclic AMP levels. These actions can lead to cell calcium overload, oxygen wasting, and arrhythmia generation.

Basic mechanisms of GIK protection

Shift from FFA to glucose uptake and utilization

The “Rackley” GIK regimen was developed to maximize myocardial glucose uptake and decrease arterial FFA levels, myocardial FFA uptake, and FFA toxicity.¹⁵ It consists of a solution of 30% glucose (300 gm/L), 50 units of regular insulin per liter, and 80 mEq of KCl/L, given intravenously at 1.5 ml/kg/hour, or approximately 100 ml per hour for a 70 kg patient. This regimen increases myocardial glucose uptake by 250% and reduces FFA uptake by 90%. The substrate shift from FFA to glucose has several beneficial effects.

- Better oxygen-efficient oxidative ATP synthesis with glucose: oxidative ATP synthesis is 11% more oxygen-efficient for glucose than for FFA oxidation;¹⁶ ie, the oxidative synthesis of equivalent amounts of ATP requires 11% more oxygen if FFA is the substrate.

- Anti-FFA actions of glucose: GIK has important anti-FFA effects.¹¹ High circulating levels of glucose and insulin both depress plasma levels of FFA and decrease myocardial FFA uptake at any given plasma FFA level,^{11,12} thereby minimizing intracellular FFA toxicity. Consequently, high glucose and insulin levels minimize the inhibition of glycolysis by FFA.

- Increased glycolytic ATP synthesis and cellular energy status: In addition to more efficient oxidative ATP synthesis, high glucose (G) and insulin (I) levels during low-flow ischemia also increase glycolytic ATP synthesis, thereby improving the high energy phosphate status of the myocardium by maintaining higher ATP and phosphocreatine levels, and lower inorganic phosphate (Pi), and ADP levels. The combination of a higher [ATP] and lower [Pi] ADP results in a significantly higher calculated free energy yield from ATP hydrolysis. Thus, the increase in ischemic glycolytic ATP synthesis acts as a “trap” for inorganic phosphate and ADP with a resulting amplification of free energy yield, which is available to all cellular ATPase reactions.⁶⁻⁸

- Effects on glycogen: Glucose and insulin preserve and restore myocardial glycogen stores. The clinical importance of these changes is suggested by the positive correlation between enhanced glucose uptake, glycogen stores, and contractile function in patients undergoing revascularization for coronary artery disease.²¹

- Effects on ion homeostasis: Glycolytic ATP protects membranes, drives the transport of Ca^{2+} into the sarcoplasmic reticulum, improves sodium homeostasis of ischemic myocardium, and regulates ATP-sensitive K^+ channels.²²⁻²⁶ A high glucose substrate increases myocyte resistance to the toxic effects of the increase in cell calcium concentration that occurs during hypoxia.²⁷

- Benefits during reperfusion: GIK may also improve cardiac function and efficiency during reperfusion by several mechanisms. GIK can accelerate the replenishing of depleted levels of Krebs cycle intermediates, a process called anaplerosis.^{1,10,28,29,32,33} An increased glycolytic flux generates more pyruvate which, in turn, generates Krebs cycle intermediates.

The glycolytic pathway itself may also be important in ischemia-reperfusion transitions. In isolated working hearts that were made ischemic and then reperfused, glycolysis was a highly adaptive emergency mechanism that prevented deleterious myocyte de-energization during the ischemia-reperfusion transitions.³⁴

GIK may slow the rate of development of ischemic necrosis; this would expand the time during which reperfusion therapy can salvage ischemic myocardium and increase the amount of salvage that is achieved. This mechanism is suggested by experimental studies showing that the protective effects of a high glucose and insulin substrate were much greater in concert with reperfusion than during the ischemic period itself.⁶ Moreover, in the Estudios Cardiologicos Latinamerica (ECLA) clinical trial (see below), GIK reduced the AMI mortality rate in patients who also received reperfusion therapy, but GIK did not reduce mortality in the absence of reperfusion therapy. Also, experimental studies have shown that GIK prevents the increase in coronary resistance that occurs with reperfusion after severe ischemia⁶ (“no reflow” phenomenon) and may, therefore, increase the successful achievement of reperfusion itself.

The insulin component of GIK may also protect the myocardium during reperfusion by activating “innate cell survival pathways” such as Akt.^{30,31}

Increased potential of GIK in the hypertrophied and/or chronically infarcted heart

An acute MI often occurs in patients with cardiac hypertrophy from chronic hypertension and/or prior chronic infarction. Hearts with pressure-overload hypertrophy are more sensitive to ischemic injury³⁵⁻³⁹ and derive greater anti-ischemic protection from increased glycolytic substrate than non-hypertrophied hearts.^{35,40} In hearts with chronic infarction, the energy reserve of the non-infarcted myocardium

is substantially impaired, increasing susceptibility to acute metabolic stress.^{41,42} Myocardial metabolic support with GIK may be particularly beneficial in these settings.

Role of insulin

Insulin is a critical component of the GIK regimen. The combination of glucose and insulin is more effective than either alone in stimulating glycolysis under ischemic conditions.⁶ Insulin may also stimulate pyruvate dehydrogenase and increase pyruvate entry into the citric acid cycle,⁴³ increase glycogen synthesis, and have direct ionic and inotropic effects.^{44,47} In studies of low-flow ischemia, insulin improved contractile function and myocardial metabolic efficiency without alterations of ATP, PCR, or Pi levels.⁴⁸⁻⁴⁹ Furthermore, in a model of ischemia and reperfusion, the inotropic and metabolic effects of insulin and epinephrine were additive and resulted in improved functional recovery in association with enhanced glucose uptake and utilization.⁵⁰ Insulin may also protect the myocardium by activating “innate cell survival pathways” such as Akt.^{30,31} In clinical practice, however, administration of insulin without glucose in non-diabetics can cause hypoglycemia. Therefore, combined glucose and insulin administration is the practical clinical regimen.

Metabolic support for the acutely loaded non-infarct region

An acute MI mechanically overloads the non-infarct region of the ventricle in proportion to MI size and the functional response of the non-infarct region may determine whether heart failure occurs. Normal myocardium is metabolically limited in its ability to adapt to an acute mechanical overload. In a study of aortic banding in mice, wild-type animals had a decreased high-energy phosphate profile, developed rapid LV failure, and had a mortality rate of 40 % at 8 weeks.⁵¹ In contrast, transgenic mice with a cardiac-specific over-expression of the glucose transporter, GLUT-1, had a markedly increased level of glucose uptake, maintained a normal high energy phosphate profile and normal left ventricular (LV) function, and had a very low mortality rate. Thus, enhanced cellular glucose uptake appears critical for the function and survival of acutely overloaded myocardium. By increasing glucose availability and uptake, GIK may provide important metabolic support to the acutely overloaded non-infarct region.

Role of residual blood flow for efficacy of GIK

With very severe ischemia, glycolytic stimulation by GIK could potentially increase myocardial acidosis from lactate accumulation. Approximately 10% of normal myocardial perfusion is required to prevent severe tissue lactate accumulation. When coronary flow was reduced by 90%, GIK doubled the glycolytic flux, prevented contracture during ischemia and reperfusion, increased post-ischemic contractile function, decreased ultrastructural damage, and increased high energy phosphate levels (Figure 2). Since the acute infarct region retained >10% of normal perfusion in most cases of acute MI, it appears to be above the perfusion level required for adequate lactate washout and successful stimulation with GIK.^{2,5} Importantly, even with more severe degrees of ischemia, although a high G+I substrate was ineffective in affording protection in some studies, it did not cause more injury than the control glucose levels.^{6,17,18}

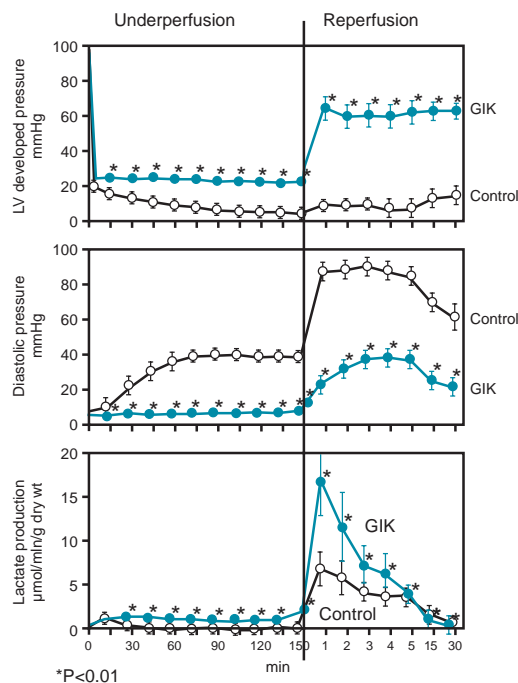
Clinical use of GIK for AMI

Limitations of early trials

Early clinical trials of GIK for acute MI yielded conflicting and inconclusive results because of several factors (eg, inadequate sample size, poor design, methodological differences including varying times to the start of GIK and the use of different GIK regimens). Many trials continued recruitment longer than 48 hours after the onset of chest pain, a timing too late to influence MI size. Several trials used oral glucose therapy with subcutaneous insulin injections or inadequate amounts of glucose, and did not achieve the glucose and insulin plasma concentrations required to significantly decrease plasma FFA concentrations. Moreover, none of these early trials had an adequate

Figure 2: Effect of GIK in simulated MI with reperfusion.⁶

The figure reports results from isolated blood-perfused isovolumic (balloon-in-LV) rabbit heart experiments. The entire LV was subjected to a 90% coronary flow reduction for 2.5 hours and then reperfused. Normal levels of glucose and insulin (control) were compared to high levels (GIK). The upper panel shows that the GIK group maintained higher levels of systolic function during ischemia, and recovered substantially better during reperfusion. The middle panel shows that during ischemia the control hearts had marked diastolic dysfunction and this became very severe with reperfusion; the GIK hearts had no diastolic dysfunction during ischemia, and milder, transient diastolic dysfunction with reperfusion. The bottom panel reports lactate production as assessed by measurement of coronary venous blood. In the GIK group, the higher levels of lactate indicated that the GIK substantially increased glycolytic flux relative to controls and that significant lactate washout occurred despite the 90% flow reduction. During reperfusion, a substantial amount of tissue lactate was washed out; despite this lactate accumulation, function was improved during both ischemia and reperfusion by the GIK perfusate.



sample size and, therefore, lacked the statistical power to rigorously assess GIK. It is not surprising that such studies showed inconsistent results.⁵²

Pre-thrombolytic era trials

A 1997 review identified 9 GIK placebo-controlled mortality trials for acute MI in which GIK (or placebo) was started within 48 hours of chest pain (Table 1).⁵² Patients in these trials did not receive reperfusion therapy (thrombolysis or percutaneous coronary intervention [PCI]). In these trials involving 1932 patients, GIK significantly reduced in-hospital MI mortality (16% versus 21% for placebo). The number of lives saved was 49 per 1000 patients treated. In the 4 studies in which GIK was administered at high concentrations (see the “Rackley GIK regimen” above¹³), the relative reduction in in-hospital mortality was even greater (6.5% versus 12%). The review concluded that GIK was highly likely to reduce mortality risk in acute MI.

Benefits of GIK with reperfusion therapy for acute MI

Three major, randomized, placebo or “standard care” controlled trials of GIK for acute MI have been conducted in concert with reperfusion therapy, with either thrombolytic drugs or percutaneous transluminal coronary angioplasty (PTCA): the DIGAMI, ECLA, and Dutch studies. All have shown impressive benefits of GIK treatment for acute MI, but none are conclusive enough to provide a definitive recommendation for the use of GIK.

Table 1: Summary of the major clinical trials of GIK for AMI

Study/clinical condition	Sample size	Mortality decrease (subgroup benefit)
GIK meta-analysis ⁵²	1932	28% (P=0.004)
DIGAMI ^{53,54}	620	58% (P=0.05; Low risk subgroup only)
ECLA ^{55,56}	252	66% (P=0.004; Reperfusion subgrp. only)
Post-op cardiogenic shock ⁶⁹	322	34% (P<0.02)
Dutch trial ⁵⁸	940	72% (P<0.05; Killip 1 + PTCA only)

*NSTEMI+STEMI **STEMI only AMI = acute myocardial infarction

The DIGAMI study

In the DIGAMI (Diabetes Mellitus, Insulin Glucose Infusion in Acute Myocardial Infarction) trial, 620 patients with diabetes and acute MI were randomly assigned to intensive insulin therapy (insulin-glucose infusion for 24 hours followed by subcutaneous insulin 4 times daily for ≥ 3 months) or standard treatment.^{53,54} Therapy was initiated at an average of 7 hours after symptom onset. Approximately half the patients received thrombolytic therapy and the treatment and control groups had similar rates of revascularization procedures.

In the intensive insulin group, there was a significant reduction in mortality at 1 year (19% vs. 26%) and at 3.4 years (33% vs. 44%). The benefit of intensive therapy was most pronounced in patients without prior insulin use and at low cardiovascular risk. In this subgroup, in-hospital mortality was significantly reduced (5% versus 12% for standard therapy).⁵⁴ Thus, the DIGAMI results indicate that an insulin-glucose infusion can significantly reduce acute MI mortality in diabetics; this may reflect the benefit of improved post-MI glycemic control, as well as the acute administration of insulin and glucose.

The ECLA study

The ECLA (Estudios Cardiológicos Latinoamerica) group randomly assigned 407 patients to high- or low-dose GIK or placebo; 62% received a thrombolytic agent.^{55,56} The ECLA study is summarized in Figure 3. For the entire group, GIK therapy was associated with a trend towards reduced in-hospital mortality (6.7% vs. 11.5% for placebo) and a trend towards a lower incidence of CHF in the GIK group. Among the patients treated with a thrombolytic agent, GIK markedly and significantly reduced in-hospital mortality (5.2% vs. 15.2%). Reperused patients who received high-dose GIK had a significant 37% reduction in mortality at 1 year follow-up. Adverse effects of GIK treatment were minimal: 17% developed mild phlebitis and only 2% developed severe phlebitis, even though the majority of patients received the infusion via a peripheral intravenous catheter.

In the ECLA study, patients treated with thrombolytics alone (without GIK) had a relatively high mortality risk of 15.2%, slightly more than twice that of many recent large trials of thrombolysis for MI.⁵⁷ Furthermore, patients not treated with thrombolytics or GIK had a mortality risk of only 6.7%. This finding of a higher mortality risk with reperfusion is not consistent with numerous randomized trials of thrombolytic therapy for acute MI. Probable explanations for this unusual result include small subgroup size; the fact that the selection of patients for reperfusion therapy was not randomized, but left to the physician's discretion, it is therefore likely that sicker patients comprised the reperfusion group; and the relatively long time from symptom-onset to treatment initiation (average =11 hours) that may have contributed to the relatively high mortality rate in the non-GIK groups. Nonetheless, the relatively high

Figure 3: Summary of the ECLA study of GIK for AMI^{55,56}

	In-hospital mortality			RR; P
	N	Control	GIK	
Reperfusion	252	15.2%	5.2%	.34; <0.004
No reperfusion	155	6.7%	9.5%	1.41; NS

• Prospective randomized trial of GIK vs. control
• Symptoms to treatment: 11 hours; STEMI and NSTEMI
• Conclusion: GIK reduces mortality by 66% in combination with reperfusion.
• CHF: beneficial trend (28% decrease; P=NS)
• Infusion rate: 1.5 mL/kg/hr; approx. 100 mL/hr

mortality rate in the non-GIK reperused group argues for caution before accepting the dramatic mortality reduction in the GIK group.

The Dutch study

The Dutch study was the largest prospectively randomized trial of GIK treatment for acute MI and the first to be done in concert with rapid, successful primary PCI (Table 2).⁵⁸ The 940 acute MI patients were randomly assigned to GIK infusion for 8-12 hours or no infusion. For the entire patient group, the primary endpoint of 30-day mortality was not significantly reduced by GIK (4.8% versus 5.8% for the control group). However, among the 856 patients who presented without signs of heart failure (HF), GIK reduced mortality by 70%, (1.2% vs. 4.2%, P<0.01); (Table 3). In diabetics, there was a non-significant trend towards reduced mortality with GIK.

GIK was not beneficial in patients who presented with HF, possibly because the GIK was infused at a relatively high rate (twice as high as in the ECLA study) and may have caused volume overload. In the 4% of patients who presented with mild HF (Killip class 2), GIK treatment was neither beneficial, nor harmful. In the 5% who presented with severe HF (Killip classes 3 and 4), the mortality risk was higher in the GIK group than in the control group. However, the small sample sizes preclude reaching definite, statistically significant, conclusions about the effects of GIK in MI patients with HF.

Comparison of the Dutch and ECLA studies

In both the ECLA and Dutch trials, more than 85% of the acute MI patients presented without signs of HF. In these non-HF patients, GIK in combination with reperfusion markedly reduced mortality risk in both studies, by 70% (P<0.01) and 66% (P<0.004) in the Dutch and ECLA studies, respectively.

However, the interaction between HF and GIK were not concordant in the ECLA and Dutch studies. In the ECLA study, there was a non-significant trend towards a lower incidence of HF in patients who received GIK. An important treatment difference and possible explanation of the different results between these two studies was the intravenous rate of volume loading that was twice as high in the Dutch study as in the ECLA study (3 vs 1.5 mL/kg/hr). Thus, any beneficial metabolic effects of GIK in the HF patients in the Dutch study may have been outweighed by the high level of volume loading, whereas the lower rate of volume loading in the ECLA study allowed the metabolic benefits of GIK to manifest.

In the Dutch study, the potential for GIK to influence mortality risk in advanced HF was also limited by the relatively low

Table 2: Summary of the Dutch study of GIK for AMI⁵⁸

• Prospectively randomized GIK vs. Control
• N = 940; STEMI only; primary angioplasty
• Largest GIK for AMI trial to date
• Symptoms to admission: 150 min.
• Door to balloon: 45 min.

Table 3: Dutch study of GIK for AMI – 30-day mortality rates according to Killip class.⁵⁸

	N	30-day mortality		RR; P
		Control	GIK	
Killip 1	856	4.2%	1.2%	0.29; <0.01
Killip 2	38	14.3%	12.5%	0.87; NS
Killip 3	28	28.6%	50.0%	1.75; NS
Killip 4	18	50.0%	66.7%	1.33; NS

Conclusion: In the absence of heart failure (Killip 1), a category that included 90% of the patients, GIK reduced AMI mortality by 71%

- No such benefit observed in the presence of CHF
- Infusion rate: 3.0 mL/kg/hr; approx. 200 mL/hr

rate of successful reperfusion in these patients. Only 50% of Killip class 3 and 4 cases had successful reperfusion. The ECLA results suggest that GIK is beneficial only in concert with reperfusion.⁵⁵ In the entire Dutch study of 940 patients, only 23 Killip class 3 and 4 patients had successful reperfusion; such a small sample precludes any definite conclusions regarding the effects of GIK in acute MI in patients with HF.

Texas Heart Institute experience

There is also a discrepancy between the Dutch study's HF results and the Texas Heart Institute results where 322 consecutive patients with refractory HF immediately post-cardiac surgery were randomly treated with standard care or standard care plus GIK.⁵⁹ The addition of GIK to the standard regimen of inotropic drugs and intra-aortic balloon counterpulsation was associated with a significant reduction in in-hospital mortality (17.6% vs. 26.6%); the infusion rates were 0.5 to 1.0 mL/kg/hr, maximally, only one-third of the rate used in the Dutch study. In an experimental model of cardiogenic shock induced by multiple coronary occlusions, GIK substantially increased short-term survival,⁶⁰ consistent with the Texas Heart Institute results.

Considered together, these studies suggest that GIK may be potentially beneficial in MI and post-cardiac surgery patients with HF and/or shock, but that the metabolic benefit of GIK can be outweighed by excessive volume loading. In the presence of HF or shock, a reasonable strategy might be to employ a more concentrated GIK solution. Placement of a central line and careful hemodynamic monitoring would be advisable in such cases.

Timing and duration of GIK

The different timeframes of GIK treatment in the ECLA and Dutch studies raise some interesting clinical issues. In the ECLA study, the time from onset of symptoms to hospital admission was 10 to 11 hours compared to only 2.5 hours in the Dutch study. Thus, in these 2 studies, GIK reduced AMI mortality risk similarly in non-HF cases, despite a wide range of duration of ischemia prior to treatment.

Furthermore, in both studies, GIK had an impressive protective effect despite being administered for a relatively small fraction of the ischemic pre-reperfusion period. For example, in the ECLA study, ischemia had been present for 10 to 11 hours prior to hospital admission. Assuming that thrombolysis was given promptly and resulted in reperfusion within 2 hours of arrival at hospital, GIK was infused for only approximately 15% (2 of 13 hours) of the pre-reperfusion ischemic time. Similarly, in the Dutch study, ischemia had been present for 2.5 hours prior to admission, and the admission-to-PCI time was approximately 45 minutes. Assuming GIK was infused for 30 of those 45 minutes, GIK would have been given for only 15% (30 of 195 minutes) of the ischemic pre-reperfusion period.

The fact that GIK was effective despite being administered for only a relatively brief period prior to reperfusion suggests several possibilities. GIK may protect against "reperfusion injury" or provide important metabolic support during reperfu-

sion and/or GIK may be able to reverse some of the ischemic injury that occurred prior to its administration.

Summary and conclusions

The results currently available from clinical trials do not provide conclusive evidence of benefit of GIK therapy in all patients with acute MI. The major studies (the Dutch and ECLA trials) were relatively small and, although a statistically significant reduction in mortality occurred in a large subgroup in each study, it did not occur in the total population studied. In the ECLA study, GIK reduced AMI mortality significantly in the subgroup that received concomitant reperfusion treatment. In the Dutch study, where 90% of the patients had emergent reperfusion via primary angioplasty, the statistically significant mortality reduction occurred in the non-CHF subgroup. Larger trials designed to evaluate these specific subgroups are necessary before GIK therapy can be recommended.

Whether GIK is beneficial in all patients with acute MI is a critical issue to resolve. Approximately 1.1 million MIs occur each year in the United States. The Dutch results suggest that approximately one million present initially without CHF and that GIK has the potential to reduce their absolute mortality risk by 3%, therefore, saving approximately 30,000 lives each year.

Whether GIK is beneficial in acute MI patients with HF and/or shock is an equally critical issue to resolve. Although such patients comprise a relatively small percentage of the total MI population, they have the highest mortality risk and are also the least tolerant of a large volume infusion. Extrapolation from the Dutch study suggests that approximately 99,000 MIs of Killip class ≥ 2 occur annually in the U.S. with a mortality risk of 26.5%, resulting in 26,235 deaths per year, even if reperfusion is rapidly available.

References

1. Taegtmeier H. Energy metabolism of the heart: From basic concepts to clinical applications. *Curr Probl Cardiol* 1994;19:59.
2. Milavetz JJ, Giebel DW, Christian TF, Schwartz RS, Holmes DR, Gibbons RJ. Time to therapy and salvage in myocardial infarction. *J Am Coll Cardiol* 1998;31:1246.
3. Christian TF, O'Connor MK, Schwartz RS, Shepherd FJ, Gibbons RJ, Ritman EL. Technetium-99m MIBI to assess coronary collateral flow during acute myocardial infarction in two closed chest animal models. *J Nucl Med* 1997;38:1840.
4. Sabia PJ, Powers ER, Ragosta M, et al. An association between collateral blood flow and myocardial viability in patients with recent myocardial infarction. *N Engl J Med* 1992; 327:1825.
5. Chareonthaitawee P, Christian TF, O'Connor MK, et al. Noninvasive prediction of residual blood flow within the risk area during acute myocardial infarction; a multicenter validation study of patients undergoing direct coronary angioplasty. *Am Heart J* 1997; 134:639.
6. Eberli FR, Weinberg EO, Grice WN, Horowitz GL, Apstein CS. Protective effect of increased glycolytic substrate against systolic and diastolic dysfunction and increased coronary resistance from prolonged global underperfusion and reperfusion in isolated rabbit hearts perfused with erythrocyte suspensions. *Circ Res* 1991; 68:466.
7. Cave AC, Ingwall JS, Friedrich J, et al. ATP synthesis during low-flow ischemia: influence of increased glycolytic substrate. *Circulation* 2000;101: 2090.
8. Saupé KW, Eberli FR, Ingwall JS, Apstein CS. Energetic and functional responses of hibernating hearts to inotropic stimulation and increased glycolytic substrates. *Circulation* 1997;96:1-627.
9. Chin ER, Allen DG. Effects of reduced muscle glycogen concentration on force, Ca²⁺ release and contractile protein function in intact mouse skeletal muscle. *J Physiol (Lond)* 1997;498 (Pt 1):17.
10. Taegtmeier H, Goodwin GW, Doenst T, Frazier OH. Substrate metabolism as a determinant for postischemic functional recovery of the heart. *Am J Cardiol* 1997;80:3A-10A.
- 10a. Oliver MF, Kurien VA, Greenwood TW. Relation between serum-free-fatty acids and arrhythmias and death after acute myocardial infarction. *Lancet* 1968;1:710-14.
11. Oliver MF, Opie LH. Effects of glucose and fatty acids on myocardial ischemia and arrhythmias. *Lancet* 1994;343:155.
12. Opie LH. Metabolism of free fatty acids, glucose and catecholamines in acute myocardial infarction. Relation to myocardial ischemia and infarct size. *Am J Cardiol* 1975; 36:938.
13. Neely JR, Morgan HE. Relationship between carbohydrate and lipid metabolism and the energy balance of heart muscle. *Ann Rev Physiol* 1974;36:413.
14. Liedtke AJ. Lipid burden in ischemic myocardium. *J Mol Cell Cardiol* 1988; 20(suppl II):65-74.
15. Rackley CE, Russell RO, Rogers WJ, Mantle JA, McDaniel HG, Papapietro SE. Clinical experience with glucose-insulin-potassium therapy in acute myocardial infarction. *Am Heart J* 1981;102:1038-1049.

16. Opie LH. *The Heart: Physiology from Cell to Circulation*. 3rd Ed. Philadelphia.; Lippincott-Raven Publishers;1998:304.
17. Apstein CS, Gravano FN, Haudenschild CC. Determinants of a protective effect of glucose and insulin on the ischemic myocardium: Effects on contractile function, diastolic compliance, metabolism and ultrastructure during ischemia and reperfusion. *Circ Res* 1983;52:515-526.
18. King L, Boucher F, Opie LH. Coronary flow and glucose delivery as determinants of contracture in the ischemic myocardium. *J Mol Cell Cardiol* 1995;27:701-725.
19. Oldfield GS, Commerford PJ, Opie LH. Effects of preoperative glucose-insulin-potassium on myocardial glycogen levels and on complications of mitral valve replacement. *J Thorac Cardiovasc Surg* 1986;91:874-878.
20. Lolley DM, Ray JF 3rd, Myers WO, Sautter RD, Tewksbury DA. Importance of preoperative myocardial glycogen levels in human cardiac preservation. Preliminary report. *J Thorac Cardiovasc Surg* 1979;78:678-687.
21. Depre C, Vanoverschelde JL, Melin JA, et al. Structural and metabolic correlates of the reversibility of chronic left ventricular ischemic dysfunction in humans. *Am J Physiol* 1995; 268(Heart Circ Physiol 37):H1264-H1275.
22. Xu KY, Zweier JL, Becker LC. Functional coupling between glycolysis and sarcoplasmic reticulum Ca²⁺ transport. *Circ Res* 1995;77:88-97.
23. Weiss JN, Lamp ST. Glycolysis preferentially inhibits ATP-sensitive K⁺ channels in isolated guinea pig cardiac myocytes. *Science* 1987;238:67-69.
24. Jeremy RW, Koretsune Y, Marban E., Becker LC. Relation between glycolysis and calcium homeostasis in post-ischemia myocardium. *Circ Res* 1992;70:1180-90.
25. Cross HR, Radka GK, Clark K. The role of Na⁺/Ca²⁺-ATPase activity during low-flow ischemia in preventing myocardial injury: a 31P, 23Na and 87Rb NMR spectroscopic study. *Magn Reson Med* 1995;34:673-685.
26. Weiss JN, Lamp ST. Cardiac ATP-sensitive K⁺ channels. Evidence for preferential regulation by glycolysis. *J Gen Physiol* 1989;94:911-35.
27. Kondo RP, Apstein CS, Eberli FR, Tillotson DL, Suter TM. Increased calcium loading and inotropy without greater cell death in hypoxic rat cardiomyocytes. *Am J Physiol* 1998;275 (Heart Circ Physiol 44):H2272-H2282.
28. Lopaschuk GD, Wambolt RB, Barr RL. An imbalance between glycolysis and glucose oxidation is a possible explanation for the detrimental effects of high levels of fatty acids during aerobic reperfusion of ischemic hearts. *J Pharmacol Exp Ther* 1993;264(1):135-44.
29. Lopaschuk GD. Alterations in fatty acid oxidation during reperfusion of the heart after myocardial ischemia. *Am J Cardiol* 1997;80(3A):11A-16A.
30. Jonassen AK, Sack MN, Mjos OD, Yellon DM. Myocardial protection by insulin at reperfusion requires early administration and is mediated via Akt and p70s6 kinase cell-survival signaling. *Circ Res* 2001;89:1191-1198.
31. Sack MN, Yellon DM. Insulin therapy as an adjunct to reperfusion after acute coronary ischemia: a proposed direct myocardial cell survival effect independent of metabolic modulation. *J Am Coll Cardiol* 2003;41:1404-1407.
32. Liu B, Clanachan AS, Schultz R, Lopaschuk GD. Cardiac efficiency is improved after ischemia by altering both the source and fate of protons. *Circ Res* 1996;79:940.
33. Mallet RT, Hartman DA, Bunger R. Glucose requirement for postischemic recovery of perfused working heart. *Eur J Biochem* 1990;188:481.
34. Schaefer S, Prussel E, Carr LJ. Requirement of glycolytic substrate for metabolic recovery during moderate low flow ischemia. *J Mol Cell Cardiol* 1995;27:2167.
35. Cunningham MJ, Apstein CS, Weinberg EO, et al. Influence of glucose and insulin on the exaggerated diastolic and systolic dysfunction of hypertrophied rat hearts during hypoxia. *Circ Res* 1990;66:406.
36. Lorell BH, Wexler LF, Momomura S, et al. The influence of pressure overload left ventricular hypertrophy on diastolic properties during hypoxia in isovolumically contracting rat hearts. *Circ Res* 1986;58:653.
37. Menasche P, Grousset C, Apstein CS, et al. Increased injury of hypertrophied myocardium with ischemic arrest: preservation with hypothermia and cardioplegia. *Am Heart J* 1985;110:1204.
38. Wexler LF, Lorell BH, Momomura S, et al. Enhanced sensitivity to hypoxia-induced diastolic dysfunction in pressure-overload left ventricular hypertrophy in the rat: role of high-energy phosphate depletion. *Circ Res* 1988;62:766.
39. Eberli FR, Apstein CS, Ngoy S, Lorell BH. Exacerbation of left ventricular ischemic diastolic dysfunction by pressure-overload hypertrophy. Modification by specific inhibition of cardiac angiotensin converting enzyme. *Circ Res* 1992;70:931.
40. Kagaya Y, Weinberg EO, Ito N, et al. Glycolytic inhibition: effects on diastolic relaxation and intracellular calcium handling in hypertrophied rat ventricular myocytes. *J Clin Invest* 1995;95:2766.
41. Neubauer S, Horn M, Naumann A, et al. Impairment of energy metabolism in intact residual myocardium of rat hearts with chronic myocardial infarction. *J Clin Invest* 1995;95:1092.
42. Friedrich J, Apstein CS, Ingwall JS. 31P nuclear magnetic resonance spectroscopic imaging of regions of remodeled myocardium in the infarcted rat heart. *Circulation* 1995;92:3527.
43. Kobayashi K, Neely JR. Effects of ischemia and reperfusion on pyruvate dehydrogenase activity in isolated rat hearts. *J Mol Cell Cardiol* 1983;15:359.
44. Moore RD. Effects of insulin upon ion transport. *Biochim Biophys Acta* 1983;737:1.
45. Gupta MP, Makino N, Khatter K, Dhalla NS. Stimulation of Na⁺-Ca²⁺ exchange in heart sarcolemma by insulin. *Life Sci* 1986;39:1077.
46. Kato M, Kako KJ. Na⁺/Ca²⁺ exchange of isolated sarcolemmal membrane: effects of insulin, oxidants and insulin deficiency. *Mol Cell Biochem* 1988;83:15.
47. Farah AE, Alousi AA. The actions of insulin on cardiac contractility. *Life Sci* 1981;29:975.
48. Tune JD, Mallet RT, Downey HF. Insulin improves cardiac contractile function and oxygen utilization efficiency during moderate ischemia without compromising myocardial energetics. *J Mol Cell Cardiol* 1998;30:2025.
49. Tune JD, Mallet RT, Downey HF. Insulin improves contractile function during moderate ischemia in canine left ventricle. *Am J Physiol* 1998;274:H1574.
50. Doenst T, Richwine RT, Bray MS, et al. Insulin improves functional and metabolic recovery of reperfused working rat heart. *Ann Thorac Surg* 1999;67:1682.
51. Liao R, Jain M, Cui L, et al. Cardiac-specific overexpression of GLUT1 prevents the development of heart failure attributable to pressure overload in mice. *Circulation* 2002; 106:2125.
52. Fath-Ordoubadi F, Beatt KJ. Glucose-insulin-potassium therapy for treatment of acute myocardial infarction: an overview of randomized placebo-controlled trials. *Circulation* 1997;96:1152.
53. Malmberg K, Ryden L, Hamsten A, et al. Effects of insulin treatment on cause-specific one-year mortality and morbidity in diabetic patients with acute myocardial infarction. DIGAMI Study Group. Diabetes Insulin-Glucose in Acute Myocardial Infarction. *Eur Heart J* 1996;17:1337.
54. Malmberg K. Prospective randomised study of intensive insulin treatment on long term survival after acute myocardial infarction in patients with diabetes mellitus. DIGAMI (Diabetes Mellitus, Insulin Glucose Infusion in Acute Myocardial Infarction) Study Group. *BMJ* 1997;314:1512.
55. Diaz R, Paolasso EA, Piegas LS, et al. Metabolic modulation of acute myocardial infarction. The ECLA (Estudios Cardiológicos Latinamerica) Collaborative Group. *Circulation* 1998;98:2227.
56. Apstein CS. Glucose-insulin-potassium for acute myocardial infarction: Remarkable results from a new prospective, randomized trial. *Circulation* 1998;98:2223.
57. An international randomized trial comparing four thrombolytic strategies for acute myocardial infarction. The GUSTO investigators. *N Engl J Med* 1993;329:673.
58. van der Horst IC, Zijlstra F, van't Hof AW, et al. Glucose-insulin-potassium infusion inpatients treated with primary angioplasty for acute myocardial infarction: the glucose-insulin-potassium study: a randomized trial. *J Am Coll Cardiol* 2003;42:784.
59. Taegtmeyer H, Goodwin GW, Doenst T, Frazier, OH. Substrate metabolism as a determinant for postschemic functional recovery of the heart. *Am J Cardiol* 1997;80:3A.
60. Coven DL, Suter TM, Eberli FR, Apstein CS. Dobutamine and glucose-insulin-potassium (GIK) improve cardiac function and survival in a randomized trial of experimental cardiogenic shock. *Circulation* 1994;90:1-480



Carl S. Apstein, M.D., is Professor of Medicine and Physiology and Director of the Cardiac Muscle Research Laboratory at the Boston University School of Medicine. He is also a Lecturer in Medicine at Harvard Medical School and an Adjunct Professor of Medicine at Tufts University School of Medicine. He served for 15 years as Chief of Cardiology of the Boston City Hospital and is now an attending cardiologist at the Boston Medical Center. He has served on the editorial boards of *Circulation*, *Circulation Research*, and the *Journal of Molecular and Cellular Cardiology*, and as a Distinguished Visiting Professor of the British Heart Association. He was elected to the American Society of Clinical Investigation and the Association of University Cardiologists. He has written extensively on myocardial energy metabolism, a career research interest that has been supported by numerous grants, including a Research Career Development Award from the National Heart, Lung, and Blood Institute (NHLBI).

Dr. Apstein has no disclosures to announce related to the enclosed CME program

Brigham and Women's Hospital,
Cardiovascular Division website:
www.heartrdoc.org

This publication is made possible by an educational grant from

Novartis Pharmaceuticals Corporation

© 2004 Brigham and Women's Hospital, Boston, Massachusetts, which is solely responsible for the contents. The opinions expressed in this publication do not necessarily reflect those of the publisher or sponsor, but rather are those of the author based on the available scientific literature. Publisher: **SNELL Medical Communication Inc.** in cooperation with Brigham and Women's Hospital, Boston, Massachusetts. TM*Cardiology Rounds* is a Trade Mark of SNELL Medical Communication Inc. All rights reserved. The administration of any therapies discussed or referred to in *Cardiology Rounds* should always be consistent with the recognized prescribing information as required by the FDA. **SNELL Medical Communication Inc.** is committed to the development of superior Continuing Medical Education.